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pH-weighted molecular MRI in human traumatic brain injury (TBI) using amine proton chemical exchange saturation transfer echoplanar imaging (CEST EPI)

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ABSTRACT

Cerebral acidosis is a consequence of secondary injury mechanisms following traumatic brain injury (TBI), including excitotoxicity and ischemia, with potentially significant clinical implications. However, there remains an unmet clinical need for technology for non-invasive, high resolution pH imaging of human TBI for studying metabolic changes following injury. The current study examined 17 patients with TBI and 20 healthy controls using amine chemical exchange saturation transfer echoplanar imaging (CEST EPI), a novel pH-weighted molecular MR imaging technique, on a clinical 3T MR scanner. Results showed significantly elevated pH-weighted image contrast (MTR_{asym} at 3 ppm) in areas of T2 hyperintensity or edema ($P < 0.0001$), and a strong negative correlation with Glasgow Coma Scale (GCS) at the time of the MRI exam ($R^2 = 0.4777$, $P = 0.0021$), Glasgow Outcome Scale - Extended (GOSE) at 6 months from injury ($R^2 = 0.5334$, $P = 0.0107$), and a non-linear correlation with the time from injury to MRI exam ($R^2 = 0.6317$, $P = 0.0004$). This evidence suggests clinical feasibility and potential value of pH-weighted amine CEST EPI as a high-resolution imaging tool for identifying tissue most at risk for long-term damage due to cerebral acidosis.

1. Introduction

Traumatic brain injury (TBI) is a major world health problem (Ghajar, 2000; Maas et al., 2008) and it is one of the leading causes of mortality and disability (Finfer and Cohen, 2001). A cascade of biological events occurs following traumatic insult to the brain leading to long-term secondary brain damage, including tissue ischemia from impaired cerebral perfusion (Coles et al., 2002; Diringer et al., 2002), amino acid excitotoxicity (Faden et al., 1989), and interstitial acidification (Hum and Traystman, 1996; Siesjo et al., 1993) from the resulting energy depletion and metabolic crisis (Vespa et al., 2005). Each of these physiologic changes have been shown to be directly or indirectly related to the degree of neuronal damage following injury and may be useful for predicting outcome. Reduced cerebral perfusion has been shown to be linked with outcome (Kirkness et al., 2005; Prabhakar et al.,

2014), while the degree of excitotoxicity has also been linked with outcome (Timofeev et al., 2011) and is the target of a variety of treatment strategies (Willis et al., 2004). Extracellular acidosis within the brain following TBI occurs due to metabolic dysfunction and results in a variety of subsequent injury mechanisms including increased permeability of calcium acid-sensing ion channels (Yermolaieva et al., 2004), further excitotoxic (McDonald et al., 1998) or oxidative (Ying et al., 1999) injury, reduced glutamate uptake by astrocytes (Swanson et al., 1995), impairment of astrocytes to maintain glucose metabolism (Swanson and Benington, 1996), and blood-brain barrier disruption (Nagy et al., 1985). Further, measures of extracellular pH, which is intimately linked with intracellular pH (Møllergaard et al., 1994), has been shown to be a strong marker of metabolic dysfunction and has been linked with increased mortality following TBI (Clausen et al., 2005; Timofeev et al., 2013; Yokota et al., 2000). Despite human and

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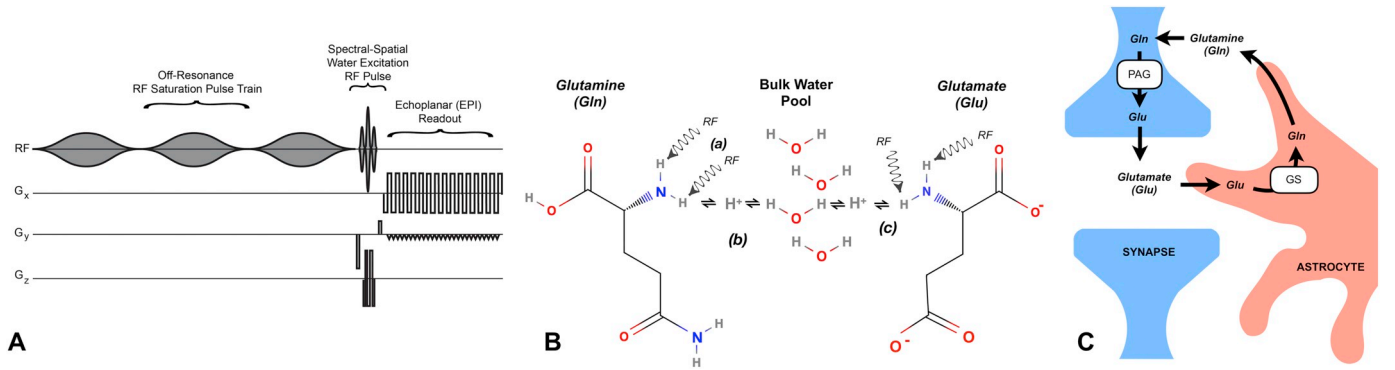


Fig. 1. A) MRI pulse sequence diagram for fast chemical Exchange saturation transfer echo planar imaging (CEST EPI) through one repetition. B) Diagram showing sites of amine proton off-resonance excitation on glutamine (a), which undergoes pH-dependent chemical exchange with the bulk water pool (b). Similarly, glutamate (c) also has exchangeable amine protons, leading to similar CEST effects. C) Diagram showing the glutamine (Gln)-glutamate (Glu) cycle. Glutamate is released from the post-synaptic neuron by either damage to the neuron or normal electrochemical activity. In functioning astrocytes, Glu is transported into the cell where it is converted to Gln via glutamine synthase (GS). Gln within the astrocytes is then shuttled to functioning neurons for Glutamate production by phosphate-activated glutaminase (PAG) or released into the extracellular space in the presence of traumatic insult to either the astrocyte or neuron.

experimental evidence suggesting acidification of the brain may occur after TBI (McIntosh et al., 1987; Shiogai et al., 1999; Zauner et al., 2002), there remains a critical need for tools to non-invasively identifying regions of the brain at risk for low extracellular pH in patients following TBI.

Amine chemical exchange saturation transfer echo planar imaging (amine CEST EPI; Fig. 1A) is a fast, noninvasive molecular MRI technique with sensitivity to amino acid concentration and pH (Harris et al., 2016; Harris et al., 2018). By preparing the MRI signal through selective saturation of the fast exchanging, pH-dependent amine protons ($-\text{NH}_2$) on glutamine or glutamate (Fig. 1B), amine CEST imaging results in signal contrast in tissues with high glutamine or glutamate concentration and low pH. Since there is a well-documented increase in both extracellular glutamate (Bullock et al., 1998; Chamoun et al., 2010; Folkersma et al., 2011; Vespa et al., 1998) and mobile glutamine (Harris et al., 2012; Palmer et al., 1993; Xu et al., 2011) following TBI from damage to neurons and astrocytes (Fig. 1C), respectively, we theorized amine CEST EPI may provide unique insight into the biologic status of the brain independent of other MR image contrasts. We hypothesized areas of T2 hyperintensity adjacent to contusion would exhibit high amine CEST EPI contrast consistent with excitotoxicity and acidification of the brain, which may be useful in subsequent studies to predicting patient outcome by more thoroughly identifying tissue most at risk for long-term damage.

2. Materials and methods

2.1. Theory

The magnetization of water protons undergoing two-compartment chemical exchange with amine protons is described by the Bloch-McConnell equations (Woessner et al., 2005) in the form of:

$$\frac{d\mathbf{M}(t)}{dt} = \mathbf{X} \cdot \mathbf{M}(t) - \mathbf{c} \quad (1)$$

where

$$\mathbf{M} = \begin{pmatrix} M_{ax} \\ M_{bx} \\ M_{ay} \\ M_{by} \\ M_{az} \\ M_{bz} \end{pmatrix}, \mathbf{X} = \begin{pmatrix} C_{2a} & k_b & -\delta a & 0 & 0 & 0 \\ k_a & C_{2b} & 0 & -\delta b & 0 & 0 \\ \delta a & 0 & C_{2a} & k_b & -\omega_1 & 0 \\ 0 & \delta b & k_a & C_{2b} & 0 & -\omega_1 \\ 0 & 0 & \omega_1 & 0 & C_{1a} & k_b \\ 0 & 0 & 0 & \omega_1 & k_a & C_{1b} \end{pmatrix}, \mathbf{c} = \begin{pmatrix} 0 \\ 0 \\ 0 \\ 0 \\ M_{a20}/T_{1a} \\ M_{b20}/T_{1b} \end{pmatrix} \quad (2)$$

pool a and pool b represent the water and amine protons, respectively; M_{a20} and M_{b20} are the equilibrium magnetizations of water and amine protons, respectively; k_b is the exchange rate of protons from pool b to

pool a ; k_a is the exchange rate of protons from pool a to pool b as given by $(M_{b0}/M_{a0}) \cdot k_b$; ω_1 is the RF pulse amplitude as given by $\omega_1 = \gamma B_1(t)$, where γ is the gyromagnetic ratio and $B_1(t)$ is given in μT ; $\delta a = \omega - \omega_a$ and $\delta b = \omega - \omega_b$, where ω is the applied radiofrequency (RF) irradiation frequency, ω_a is the water proton resonance frequency, and ω_b is the amine proton resonance frequency; T_{1a} and T_{1b} are the longitudinal relaxation times of water and amine protons, respectively; and $C_{1a} = (1/T_{1a}) + k_a$, $C_{2a} = (1/T_{2a}) + k_a$, $C_{1b} = (1/T_{1b}) + k_b$, $C_{2b} = (1/T_{2b}) + k_b$ represent the sum of exchange and relaxation rates. Eq. 1 can be solved analytically to yield

$$\mathbf{M}(t) = e^{\mathbf{X}t} \cdot \mathbf{M}_0(\mathbf{X}/\mathbf{c}) - (\mathbf{X}/\mathbf{c}) \quad (3)$$

The effects of pH on the CEST signal can then be modeled by accounting for the chemical exchange rate between amine and water protons. The chemical exchange between amine and water protons can be characterized as a base-catalyzed process, governed by the equation:

$$k_b = k_0 + k_{base} * 10^{-(14-pH)} \quad (4)$$

where k_0 is the default exchange rate, k_{base} is the base-catalyzed rate constant, and k_b is the exchange rate of protons from the metabolite proton pool to the water pool (Liepinsh and Otting, 1996).

2.2. Phantom preparation and estimation of exchange rate constants

To determine the constants k_0 and k_{base} for amino acid amines, a series of glutamine and glutamate phantoms with pH ranging from 4.8 to 7.5 were prepared for MRI evaluation. Glutamine or glutamate were combined with phosphate-buffered saline (PBS) in 50 mL falcon tubes to reach a concentration of 50 mM. 0.5 mM Gd-DTPA (Magnevist®) was then into each phantom solution to reduce relaxation times (i.e. T_1 and T_2) near physiologic levels. The pH values of sample solutions were then titrated to 18 different values ranging from 4.8 to 7.5, with acid (1 N HCl) and base (1 N NaOH) solution. Phantom pH values were re-recorded after the MRI scanning and used as the final pH values for data fitting using the Bloch-McConnell equations described previously.

2.3. Mathematical simulations of CEST effects

To demonstrate the relationship between amine CEST image contrast and pH for both glutamine and glutamate we employed the Bloch-McConnell simulation framework for CEST EPI previously described (Harris et al., 2016), using exchange rate values estimated from phantom experiments. Specifically, we simulated the z-spectrum from pH values ranging from 5.5 through 8.0 by first assuming the concentration of amine protons was 50 mM, which is reasonable

considering 1) the concentration of amino acids in normal neural tissues has been estimated around 20–25 mM (Perry et al., 1971); 2) many amino acid derivative metabolites including norepinephrine, 5-hydroxytryptophan, levodopa and other neurotransmitters possess a similar amine functional group that will also contribute to the CEST signal at 3.0 ppm; and 3) we have previously shown these amino acids exhibit similar pH-dependent exchange rate characteristics and CEST effects (Harris et al., 2015). Additionally, we assumed the MR characteristics of water protons were similar to values in normal white matter ($T_{1a} = 1.22$ s, $T_{2a} = 0.107$ s) (Harris et al., 2015; Wright et al., 2008), assumed the MR characteristics of the amine protons were similar to those in the literature ($T_{1b} = 0.2$ s and $T_{2b} = 0.1$ s) (Choi et al., 2006; Li et al., 2008; Traber et al., 2004; Woessner et al., 2005), and used a set of 3×100 ms Gaussian radiofrequency CEST saturation pulses with an amplitude of $6 \mu\text{T}$, similar to previous studies (Harris et al., 2016; Harris et al., 2018). Lastly, using similar values of T_1 and the exchange rates estimated from the phantom experiments, we simulated the CEST signal for a range of concentrations from 5 mM to 50 mM and a range of transverse relaxation times from $T_2 = 50$ ms to 500 ms.

2.4. Quantification of CEST contrast

During a CEST imaging experiment, the RF saturation frequency ω is swept across a range of values to obtain a “z-spectrum” (Bryant, 1996). To reduce the effects of T_1 and T_2 weighting along with other variables, the attenuation of bulk water magnetization following a saturation pulse is often described by the magnetization transfer ratio (*MTR*), given by:

$$MTR(\omega) = \frac{S(\omega)}{S_0} \quad (5)$$

where $S(\omega)$ is the magnitude of water proton MR signal available after the saturation pulse with frequency ω and S_0 is the MR signal available without application of any RF saturation pulses. Since *MTR* can be affected by symmetric effects of direct water saturation and conventional magnetization transfer effects, CEST contrast is often described by the asymmetry in the magnetization transfer ratio (*MTR_{asym}*) with respect to water proton resonance:

$$MTR_{asym}(\omega) = \frac{S(-\omega) - S(\omega)}{S_0} \quad (6)$$

For the current simulation, *MTR* at spectral points between -5.0 and $+5.0$ ppm were calculated to obtain simulated z-spectra in these tissues, and the *MTR_{asym}* at 3.0 ppm was used as a final measure of pH-dependent amine CEST contrast for use in subsequent patient studies.

2.5. Human participants

A total of 17 patients with traumatic brain injury and 20 healthy volunteers were enrolled in this prospective, Health Insurance Portability and Accountability Act (HIPAA) compliant study approved by the Institutional Review Board at our institution. All patients and volunteers signed informed written consent to have advanced imaging collected. A summary of TBI patient demographics is shown in Table 1.

2.6. Magnetic resonance imaging

All patients underwent anatomic and amine CEST-EPI on a 3T MR scanner (Siemens Trio, Prisma, or Skyra; Siemens Medical; Erlangen, Germany). All patients received a standardized anatomic imaging protocol consisting of at least T2-weighted turbo spin echo images, diffusion tensor images (DTI), susceptibility-weighted gradient echo images (SWI), fluid-attenuated inversion recovery (FLAIR) images, and 1 mm isotropic parameter matched 3D inversion-recovery gradient recalled echo (IR-GRE) images. Amine CEST-EPI was collected with the following scan parameters: field-of-view (FOV) = 256×256 mm,

Table 1
Traumatic Brain Injury (TBI) patient demographics.

Subject ID	Age/Gender	Injury	Diagnosis	Days from Injury to MRI	ED GCS	GCS @ MRI	6 Month GOSE
1	45M	Fall while Hang Gliding	DAI and WM Microhemorrhages	182	5	10	3
2	35M	MVA with Pedestrian	Bilateral SAH and IPH	3	7	7	3
3	28M	MVA	Bilateral hemorrhagic contusions, SAH in frontal lobe, SDH along cerebral convexities	37	11	3	4
4	37M	Fall from 25 ft. roof	Skull fracture; SDH; L. MLI and subfalcine herniation	9	4	6	2
5	20F	MVA with Pedestrian	R SDH; frontal contusions; SAH	1	3	3	3
6	74M	MVA with Pedestrian	SAH; Contusions	8	6	7	1
7	22M	MVA	L. temporal hemorrhagic contusion; SAH; SDH	1	8	10	4
8	24M	MVA with Motorcycle	Frontal extraaxial hematomas; midline shift; scattered contusions	9	6	7	7
9	22M	MVA with Bicycle	L. Frontal Hemorrhagic Contusion; diffuse microhemorrhages; DAI	18	15	14	8
10	40M	Hit in head and fall	Epidural and SDH with cerebral edema	4	3	14	5
11	49M	Fall from 15 ft	L. Frontal SDH; SAH; L. temporal and orbitofrontal hemorrhagic contusions	1	14	12	6
12	55M	MVA	Traumatic SAH; bifrontal SDH	20	3	8	1
13	59M	Fall down stairs	R SDH; SAH; hemorrhagic contusions orbitofrontal and right temporal lobe	8	3	7	1
14	50M	Bicycle accident	Bifrontal contusions, frontal SDH, scattered SAH	2	12	4	1
15	84M	Presumed fall	R frontoparietal and L. temporal contusion, SDH and SAH	2	13	7	1
16	33M	High speed motorcycle accident	Multifocal Parenchymal hemorrhagic contusion, SAH, SDH along falx	1	3	3	1
17	50F	MVA with pedestrian	temporal lobe contusion and SDH	4	14	3	1

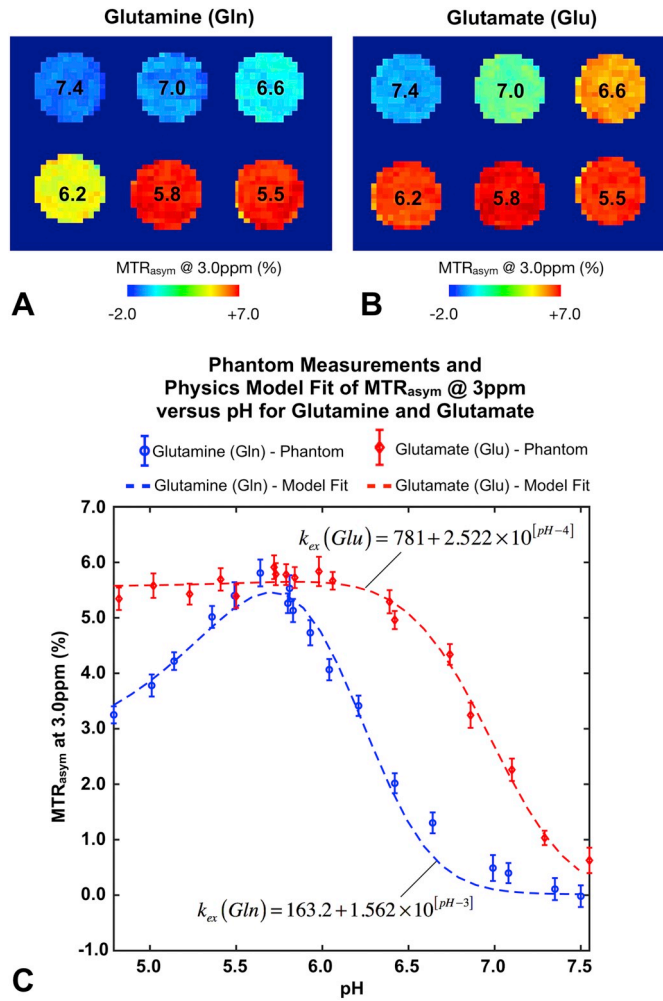


Fig. 2. Amine CEST images of A) glutamine and B) glutamate phantoms with a concentration of 50 mM and varying pH. C) Physical MRI measurements and Bloch-McConnell mathematical model fits for glutamine and glutamate at 5 mM and various pH values used to estimate pH-dependent exchange rates.

matrix size = 128×128 , number of slices = 25, slice thickness = 4 mm with no interslice gap, $TE/TR = 27/380$ ms, bandwidth = 1628 Hz, and generalized autocalibrating partially parallel acquisition (GRAPPA) factor = 2. Off-resonance saturation was applied using a pulse train of 3×100 ms Gaussian pulses with amplitude of $6 \mu T$, with spoiling gradients to remove transverse magnetization prior to each readout. A total of 25 off-resonance, or “z-spectral” points were sampled at frequency offsets of: -3.4 to -2.6 ppm, -0.3 to $+0.3$ ppm, and $+2.6$ to $+3.4$ ppm, all in increments of 0.1 ppm. Total scan time for CEST-EPI was approximately 6 min.

2.7. ΔB_0 map estimation using iterative down-sampling estimation with Lorentzian fitting

In order to estimate the static magnetic field distortion, ΔB_0 , we employed a novel method using iterative down-sampling with Lorentzian fitting (Yao et al., 2018). Briefly, all CEST data were first preprocessed with motion correction (*mcflirt* function, FSL; Oxford) (Jenkinson et al., 2002). Next, a slice-by-slice ΔB_0 map estimation was achieved by first normalizing the raw CEST data with respect to the S_0 reference scan data, resulting in z-spectral measurements, then performing k-means clustering cluster the pixels with similar patterns in the z-spectrum. The number of clusters for each iteration was set to 3^m , where m is the iteration number. Next, the average z-spectrum data was

estimated across all pixels within the same cluster, and then the mean z-spectrum data were fit to the Lorentzian function in the form: $c \frac{b}{(x-a)^2 + b^2}$, where a indicates the shift in z-spectral center point, b relates to the width of the Lorentzian shape, c and b together determine the y-scaling of the function. The constraint bound for ΔB_0 estimation was set to $\pm 1/m$ ppm. The magnetic field distortion, or inhomogeneity in the static magnetic field (ΔB_0), was estimated by the value of a in the Lorentzian model and subsequently stored for all pixels. Lastly, a 3D Gaussian filter with standard deviation of unity was applied to the final ΔB_0 map to remove potential erroneous estimations.

2.8. Region of interest selection

All images were registered to high-resolution, 1 mm isotropic T1-weighted images using a 6° of freedom rigid body transformation and mutual information cost function (FSL, FMRIB, Oxford, UK) for subsequent analyses. MTR_{asy} at 3 ppm and magnetic field fluctuations (ΔB_0) were measured within areas of T2 hyperintensity along with areas of contusion as defined by signal voids on susceptibility (T_2^*) weighted gradient echo images. Additionally, regions with apparent diffusion coefficient (ADC) $< 0.6 \mu m^2/ms$, suggestive of ischemia, were included. Measurements in patients with multiple sites of T2 hyperintensity, contusion, or potential ischemia were averaged together resulting in a single measurement per patient. As a reference, a 2 cm diameter spherical region of interest was identified within normal-appearing white matter (NAWM), distal from any apparent lesion, hemorrhage or contusion as well as a similar region of interest in white matter within the brains of healthy volunteers. Line plots ranging from 13 to 20 mm were created by traversing the edge of the T2 hyperintense lesion in TBI patients in order to document any apparent spatial colocalization of acidic and ischemic tissue using MTR_{asy} and ADC, respectively.

2.9. Clinical examination and outcome measures

The Glasgow Coma Scale (GCS) is a commonly used clinical measure of initial TBI severity, has been shown to be an early prognostic marker for outcome (Jiang et al., 2002), and is inversely correlated with likelihood of significant morbidity and mortality (Jiang et al., 2002; Lawrence F. Marshall et al., 1991). Average GCS at the time of MRI was 7.4 ± 0.9 S.E.M. (range = 3 to 14). Additionally, we evaluated the extended Glasgow outcome scale (GOSE) at 6 months from the time of injury as a widely accepted clinical measure of outcome in patients with TBI (Jiang et al., 2002; Marshall et al., 1991; Levin et al., 2001; Woischneck and Firsching, 1998). Six-month GOSE scores were available for 11 of the 17 patients in the current study and averaged 4.2 ± 0.6 S.E.M. (range = 1 to 8). In the current study we tested the hypothesis that CEST measures of MTR_{asy} at 3 ppm within T2 hyperintense areas adjacent to contused tissue correlated with GCS at the time of the MRI examination and could predict GOSE 6 months following initial injury.

3. Statistical analyses

All values are presented as mean \pm standard error of the mean (S.E.M.). A linear mixed effects model and expected marginal means estimations were used to compare measurements of MTR_{asy} at 3 ppm in regions of normal appearing white matter (NAWM), T2 hyperintense regions, areas of contusion, areas of potential ischemia as indicated by $ADC < 0.6 \mu m^2/ms$, and healthy white matter from control participants. Similarly, a linear mixed effects model and expected marginal means estimations were used to compare measurements of ΔB_0 at 3 ppm in regions of normal appearing white matter (NAWM), T2 hyperintense regions, areas of contusion, and measurements from white matter in healthy volunteers. A total of 5 comparisons were performed for MTR_{asy} and ΔB_0 at 3 ppm, so a Bonferroni-corrected p -value of

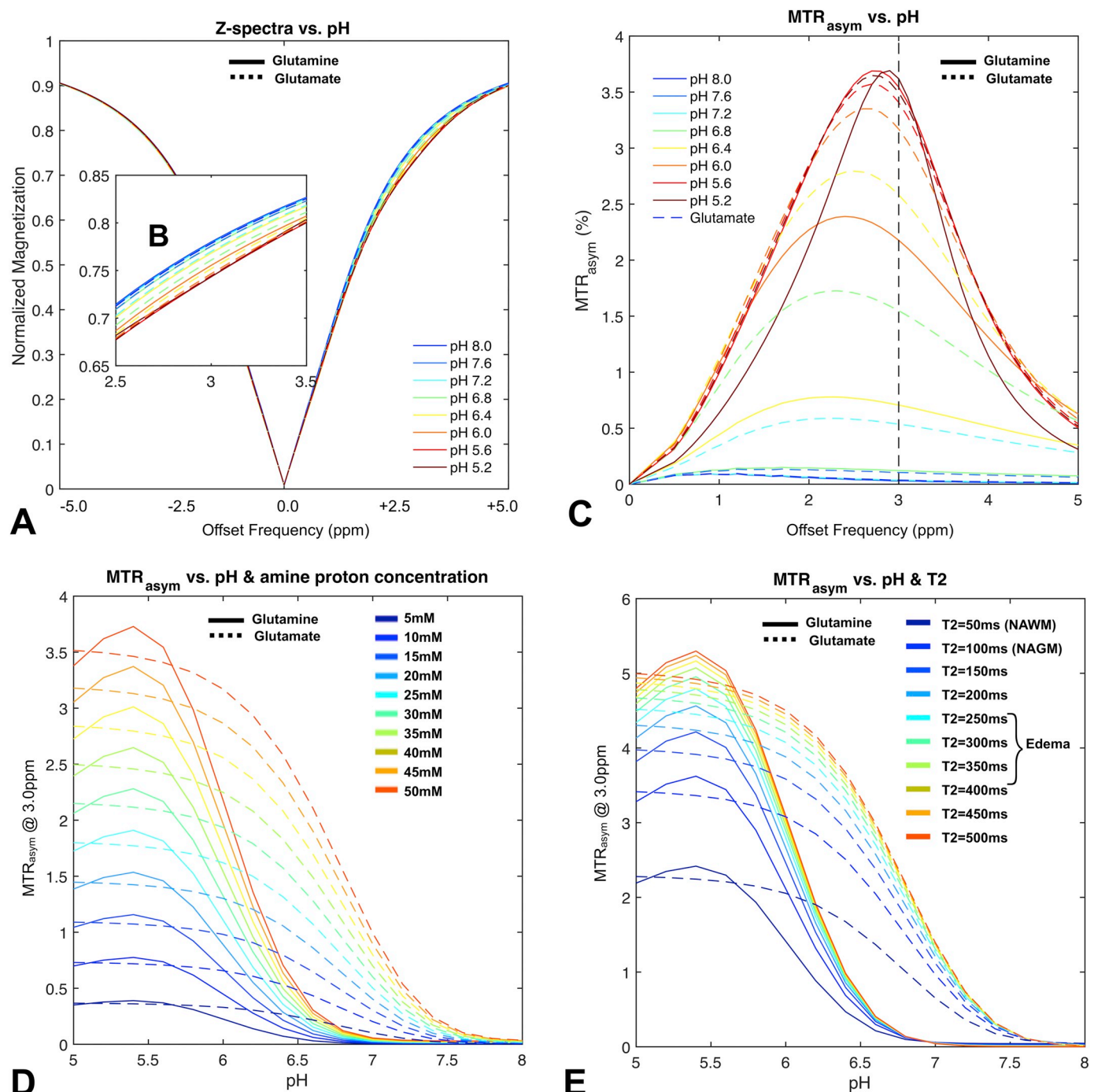


Fig. 3. A) Bloch-McConnell MR simulation results showing z-spectra for glutamine and glutamate at 50 mM concentration for various pH. B) Zoomed region of the z-spectrum showing increasing attenuation of the MR signal at offset frequencies around +3.0 ppm with decreasing pH. C) Asymmetry in the magnetization transfer ratio about water resonance (MTR_{asym}) for glutamine and glutamate at 50 mM and various pH showing maximum asymmetry around +2–3.0 ppm. D) MTR_{asym} at 3.0 ppm for glutamine and glutamate at various concentrations versus pH. E) MTR_{asym} at 3.0 ppm for glutamine and glutamate at 50 mM and various tissue T2 relaxation times versus pH.

0.01 was used to indicate statistical significance. Additionally, linear regression was used to compare CEST measures of MTR_{asym} at 3 ppm within areas of T2 hyperintensity to both GCS at the time of MRI examination and GOSE at 6 months from injury in patients with TBI. We used nonlinear regression to test whether there was a logarithmic association between the time between injury and MRI exam and MTR_{asym} at 3 ppm within areas T2 hyperintensity, which may suggest a higher degree of excitotoxicity and acidosis in the early acute phases of TBI. Lastly, we used a multiple linear regression using a combination of CEST measures and the time between injury and MRI exam in order to

predict both GCS at the time of the MRI exam and GOSE at 6 months from injury.

4. Results

4.1. Phantom and simulation results

Phantom experiments involving CEST imaging of 50 mM vials containing glutamine and glutamate exhibited distinct changes at lower pH values (Fig. 2A–B), consistent with previous studies. Results clearly showed

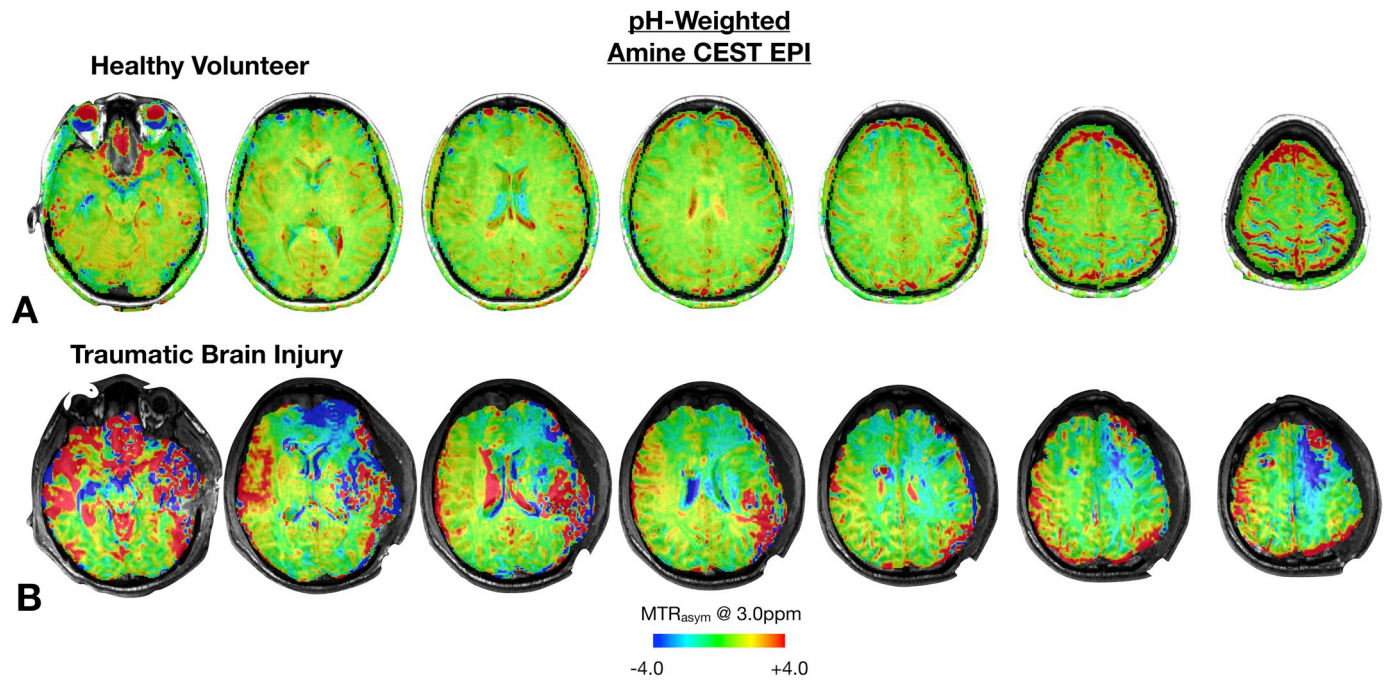


Fig. 4. Example of CEST-EPI estimates of MTR_{asym} at 3 ppm across multiple slices in a A) a 23 year old healthy volunteer and B) a 22 year old patient with TBI (Subject #7).

a higher MTR_{asym} at 3 ppm for pH values below around 7.0 in for glutamine and around 7.5 for glutamate, with an estimated pH-dependent exchange rate, in Hz, for glutamine and glutamate of $k_{\text{ex}}(\text{Gln}) = 163.2 + 1.562 \times 10^{[\text{pH}-3]}$ and $k_{\text{ex}}(\text{Glu}) = 781 + 2.522 \times 10^{[\text{pH}-4]}$, respectively (Fig. 2C).

Consistent with previous simulations and other studies (Harris et al., 2016; Harris et al., 2015; Harris et al., 2018; Jin and Kim, 2014; Jin et al., 2012; Sun et al., 2013), CEST measures of MTR_{asym} at 3 ppm from bulk water for both glutamine and glutamate demonstrated sensitivity to pH, concentration, and T2 relaxation time. In particular, the z-spectrum showed a slight decrease in normalized MR signal intensity with decreasing pH assuming a constant glutamine concentration of 50 mM (Fig. 3A–B). The MTR_{asym} around 2–3 ppm showed the optimal sensitivity to pH-related amine proton exchange (Fig. 3C), which increased with increasing glutamine and glutamate concentration (Fig. 3D) and increasing tissue T2 (Fig. 3E), which supports the hypothesis that amine CEST EPI may provide value in identifying tissue at risk of acidity due to excitotoxicity, particularly in areas of T2 hyperintensity.

4.2. MTR_{asym} at 3 ppm within NAWM, T2 hyperintensity, contusion and ischemia in TBI

Whole brain CEST-EPI data within healthy volunteers exhibited relatively uniform, low values of MTR_{asym} at 3 ppm (Fig. 4), averaging $0.58\% \pm 0.05\%$ standard error of the mean (S.E.M.), which was similar to average values of MTR_{asym} at 3 ppm in a 2 cm diameter region of relatively NAWM on anatomic MRI in TBI patients (Fig. 7A; $0.66\% \pm 0.05\%$). No difference in MTR_{asym} at 3 ppm was observed between NAWM in TBI patients and white matter in healthy volunteers ($P = 0.2371$). This was in stark contrast to MTR_{asym} at 3 ppm within lesions in TBI patients (Fig. 4). As illustrated in Fig. 5A–C and quantified in Fig. 7A, regions of T2 hyperintensity near areas of contusion exhibited a significantly higher estimate of median MTR_{asym} at 3 ppm ($1.62\% \pm 0.09\%$) compared with NAWM (Fig. 7A; $P = 0.0007$). Average values of MTR_{asym} at 3 ppm in a 2 cm diameter region of relatively NAWM on anatomic MRI in TBI patients yielded an average value of $0.66\% \pm 0.05\%$ standard error of the mean (S.E.M.) (Fig. 7A).

In many cases, the acidic lesion appeared larger and more extensive than lesions on anatomic MRI scans (Fig. 6A,B). In the 13 of 17 patients, area of diffuse axonal injury, contusion or hemorrhage on anatomic MRI (as indicated with T2*-weighted SWI) had negative MTR_{asym} at 3 ppm and were easily identifiable on individual patient scans and were often localized within area of the corpus callosum (Fig. 6A–C). Median MTR_{asym} at 3 ppm measured approximately $-0.92\% \pm 0.25\%$, which was significantly lower than NAWM and the T2 hyperintense lesion (Fig. 7A; $P < 0.0001$). Areas of potential ischemia as indicated by $ADC < 0.6 \mu\text{m}^2/\text{ms}$ exhibited an average MTR_{asym} at 3 ppm around $0.86\% \pm 0.34\%$, which was not significantly different from NAWM (Fig. 7A; $P = 0.4988$). No significant association was observed between ADC and MTR_{asym} at 3 ppm in areas of T2 hyperintensity (Fig. 7B; $R^2 = 0.1267$, $P = 0.1609$), as also illustrated by the line plots through the T2 hyperintense lesions in Fig. 5A–C, nor between ADC and MTR_{asym} at 3 ppm in areas of contusion (Fig. 7C; $R^2 = 0.1438$, $P = 0.2013$).

4.3. Magnetic field distortions (ΔB_0) within NAWM, pericontusion, and hemorrhage in TBI

Perturbations or distortions in the homogeneous, static magnetic field within the MRI system can lead to errors in MTR_{asym} measurements and may provide unique sensitivity to regions of resolving blood products. The median ΔB_0 within NAWM averaged $0.042 \text{ ppm} \pm 0.011 \text{ ppm}$ S.E.M., which was not significantly different (ANOVA, $P = 0.1287$) from the ΔB_0 estimated within areas of T2 hyperintensity (Fig. 7D; $0.039 \text{ ppm} \pm 0.018 \text{ ppm}$) or contusion (Fig. 7D; $0.077 \text{ ppm} \pm 0.012 \text{ ppm}$). No difference was observed between NAWM in TBI patients and white matter from healthy volunteers ($P = 0.8213$), which averaged $0.045 \text{ ppm} \pm 0.009 \text{ ppm}$ S.E.M..

4.4. Correlation between MTR_{asym} at 3 ppm and clinical outcome

A strong negative correlation was observed between GCS at the time of MRI and MTR_{asym} at 3 ppm within contusion plus pericontusional tissue (Fig. 8A; $R^2 = 0.4777$, $P = 0.0021$), suggesting patients with the highest degree of brain acidity had worse function. Similarly, MTR_{asym}

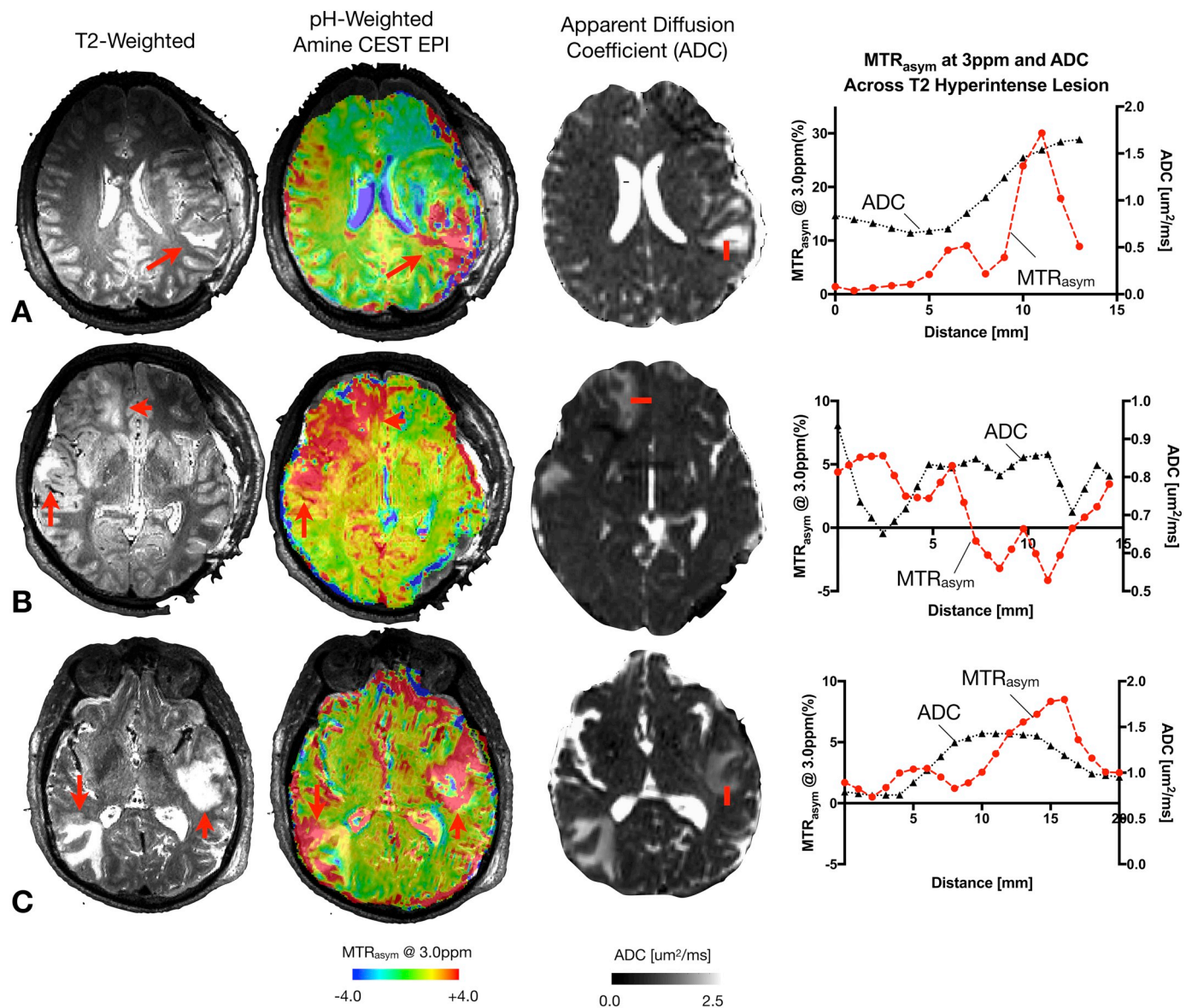


Fig. 5. T₂-weighted anatomic MRI (far left column), pH-weighted amine CEST EPI (MTR_{asym} at 3 ppm, left column), apparent diffusion coefficient (ADC) map (right column), and a line plot showing MTR_{asym} at 3 ppm and ADC through the T2 hyperintense lesion (far right column) in A) a 22 year old male patient (Subject #7) with a left temporal hemorrhagic contusion, subarachnoid hemorrhage, and subdural hematoma following being struck by a motor vehicle; B) a 24 year old male (Subject #8) with a series of frontal extra-axial hematomas, significant midline shift from cerebral edema, and scattered contusions after being hit on a motorcycle by a motor vehicle; and C) a 84 year old male patient (Subject #15) suffered a presumed fall resulting in right frontoparietal and left temporal contusions, subdural hematoma, and subarachnoid hemorrhage. Red arrows on T2-weighted images show sites of T2 hyperintense lesions. Red lines on ADC maps show where line plot measurements were obtained. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

at 3 ppm was inversely correlated with GOSE at 6 months (Fig. 8B; $R^2 = 0.5334$, $P = 0.0107$), also suggesting patients with the lowest pH had worse long-term outcomes. We also observed a significant non-linear (logarithmic) association between MTR_{asym} at 3 ppm and the time between the injury and MRI exam (Fig. 8C; Model: $\log_{10}(\text{time from injury}) = B \times [MTR_{\text{asym}}@3\text{ppm}] + C$; $R^2 = 0.6317$, $P = 0.0004$), implying a higher acidity during acute injury compared with more sub-acute or chronic stages of TBI. Lastly, multiple linear regression suggested a combination of MTR_{asym} at 3 ppm along with the time between injury and MRI could better predict GCS at the time of the MRI exam (Fig. 8D; Model: $\text{GCS} = B_1 \times \log_{10}(\text{time from injury}) + B_2 \times [MTR_{\text{asym}}@3\text{ppm}] + C$; $R^2 = 0.5796$, $P = 0.0004$) and GOSE at 6 months (Fig. 8E; Model: $\text{GOSE} = B_1 \times \log_{10}(\text{time from injury}) + B_2 \times [MTR_{\text{asym}}@3\text{ppm}] + C$; $R^2 = 0.6539$, $P = 0.0026$).

5. Discussion

Following TBI, a series of secondary brain damage occurs including reduced cerebral blood flow, hypoxia, and excitotoxicity (Bullock et al., 1995; Bullock et al., 1998; Miller, 1985), all of which result in cerebral acidosis (Clausen et al., 2005; Hovda et al., 1992; Kalimo et al., 1981). Results from the current study are consistent with the hypothesis that areas of T2 hyperintensity adjacent to contusions exhibit significant acidosis, arising from excitotoxicity. Further, our results suggest the degree of acidosis is highly correlated with both clinical function as well as outcome, as evidenced by correlation with GCS at the time of the MRI examination and GOSE at 6 months, respectively. Eloquent studies using multiparametric sensors inserted into the brain in patients with TBI have verified a potential association between low extracellular pH and outcomes including mortality (Gupta et al., 2004; Timofeev

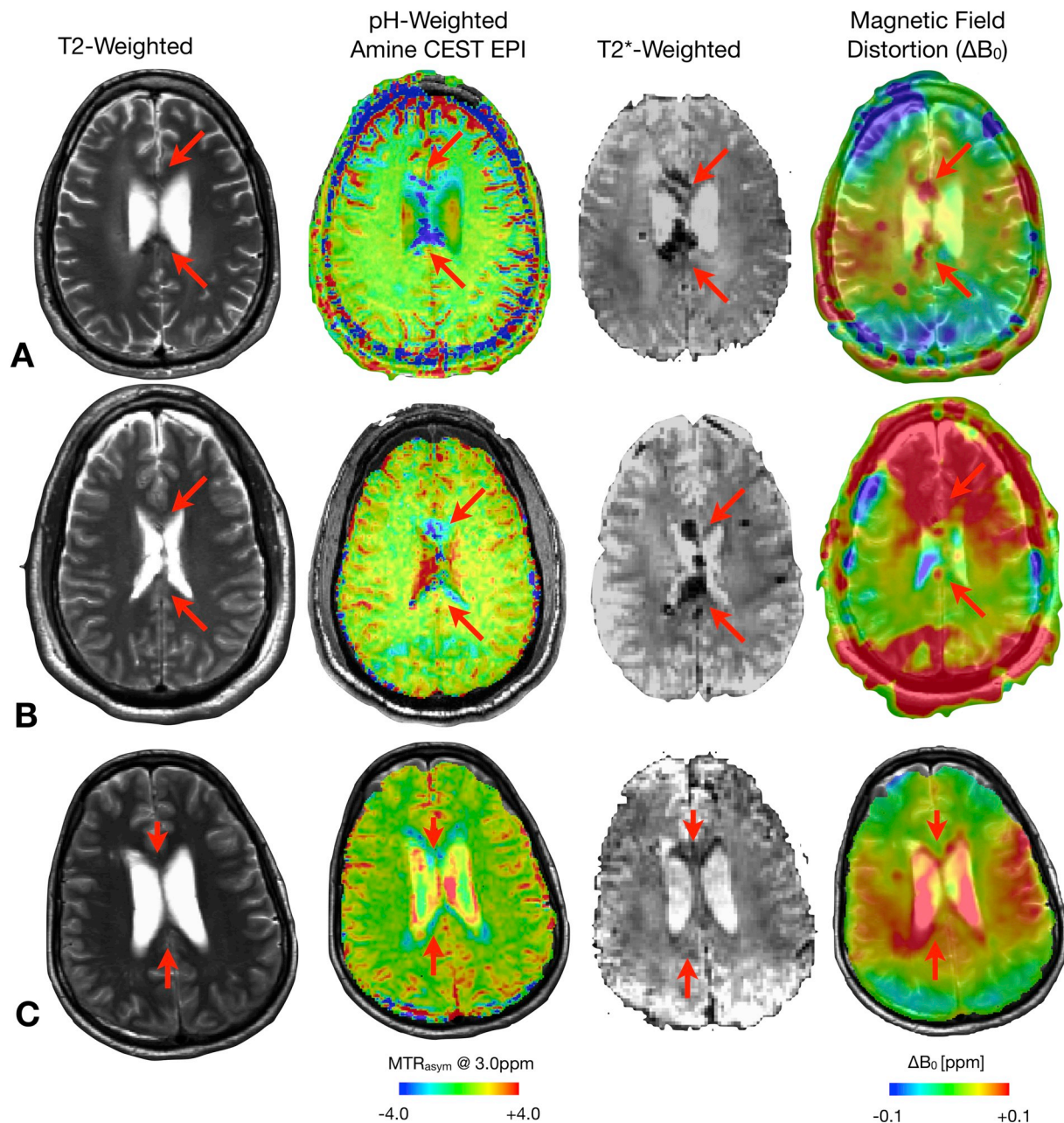


Fig. 6. T₂-weighted anatomic MRI, pH-weighted amine CEST EPI, T₂*-weighted MRI, and estimated magnetic field distortion (ΔB_0) (Left-to-Right) for A) a 45 year old male patient (Subject #1) with diffuse axonal injury and white matter microhemorrhages after suffering a fall from hang gliding; B) a 35 year old male patient (Subject #2) who was struck by a motor vehicle and suffered bilateral subarachnoid and intraparenchymal hemorrhage; and C) a 28 year old male subject (Subject #3) who suffered bilateral hemorrhagic contusions, subarachnoid hemorrhage in the frontal lobe, and subdural hematomas along the cerebral convexities after being in a motor vehicle accident. Red arrows highlight areas of microbleeds and/or diffuse axonal injury within the corpus callosum. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

et al., 2013). Interestingly, we also observed a correlation between time from initial injury and degree of acidosis as evidenced by CEST contrast, which appears to be consistent with these same studies involving temporal pH monitoring using intracranial sensors, which showed acidic pH measurements reversing or normalizing in subsequent days following injury (Timofeev et al., 2013).

Few studies have used molecular MR imaging to evaluate pH changes following TBI, and the current study represents the first to utilize amine CEST EPI on 3T clinical MR systems to evaluate acidosis in human TBI at high spatial resolution. A number of preclinical studies have shown alterations in intra- and extracellular pH using phosphorous nuclear magnetic resonance (³¹P NMR) spectroscopic techniques (McIntosh et al., 1987; Vink et al., 1987). However, ³¹P NMR has

extremely low sensitivity, therefore data must be acquired using large voxels and long scan times, making it less suitable for routine clinical examination and differentiation of acidic subregions in heterogeneous lesions. A recent study by Wang et al. (2017) utilized amide proton transfer (APT) CEST imaging on a preclinical MR system (4.7T) in untreated rats following TBI, noting a characteristic decrease in MTR_{asym} at 3.5 ppm (for amide protons), which has been shown to similarly reflect a decreasing pH (Note: decreasing pH results in increased MTR_{asym} at 3 ppm for amine protons and decreased MTR_{asym} at 3.5 ppm for amide protons (Jones et al., 2013)). Further, the approach in the current study focusing on amine CEST imaging has specific advantages over APT imaging including short radiofrequency irradiation times due to the fast exchange rates of the amine protons, so fast readout strategies including

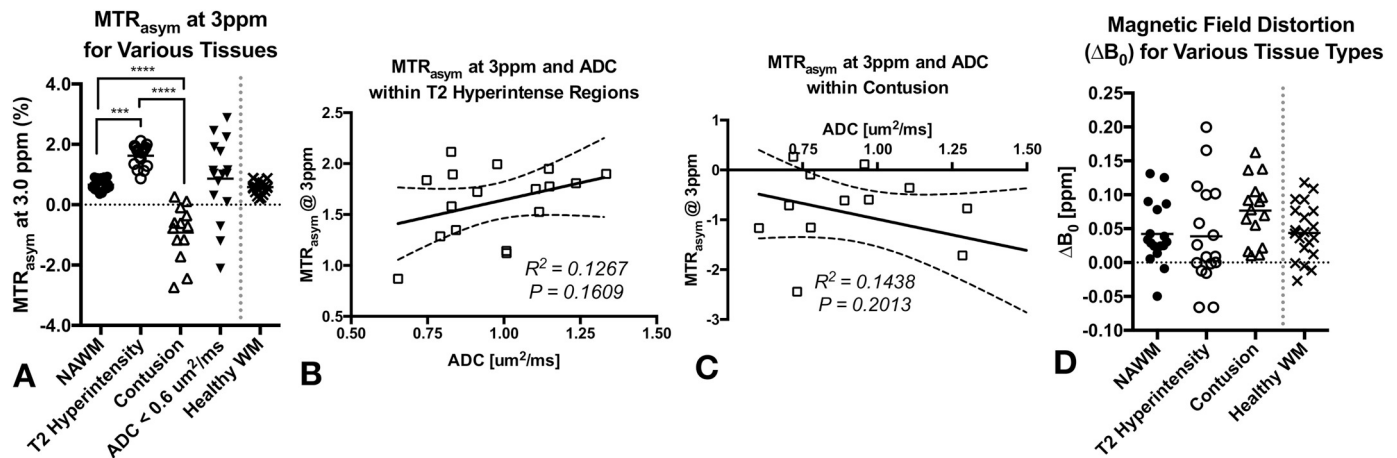


Fig. 7. A) Amine CEST EPI measures of MTR_{asym} at 3 ppm (acidity) in normal appearing white matter (NAWM, solid circles), T₂ hyperintense (open circles), regions of contusion T₂*-weighted imaging (open triangles), and regions of potential ischemia with ADC < 0.6 μm²/ms. B) Correlation between MTR_{asym} at 3 ppm and ADC within T₂ hyperintense lesions. C) Correlation between MTR_{asym} at 3 ppm and ADC in areas of contusion. D) Estimation of magnetic field distortion (ΔB₀) in normal appearing white matter (NAWM, solid circles), T₂ hyperintense (open circles), and regions of contusion T₂*-weighted imaging (open triangles). *P < 0.05; **P < 0.01; ***P < 0.001; ****P < 0.0001.

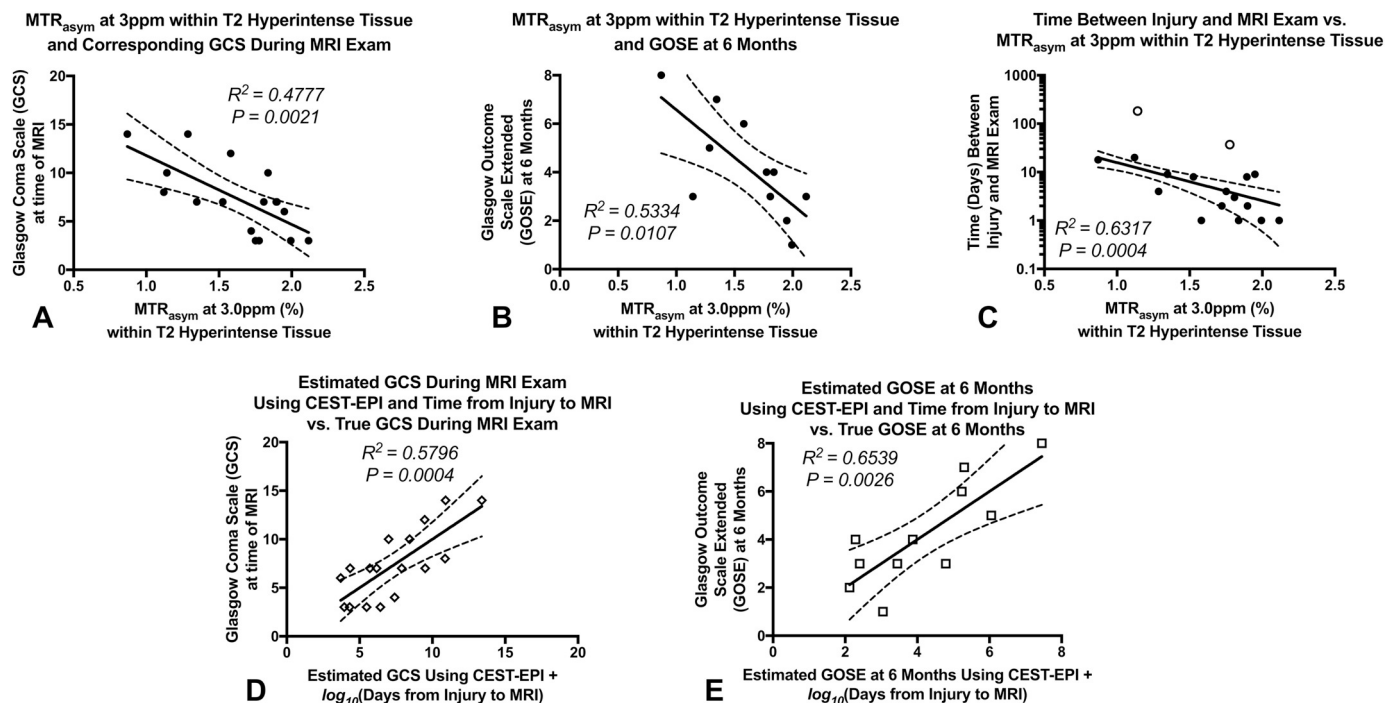


Fig. 8. A) Correlation between MTR_{asym} at 3 ppm within T₂ hyperintense regions and Glasgow Coma Scale (GCS) at the time of the MRI exam ($R^2 = 0.4777$, $P = 0.0021$). B) Correlation between MTR_{asym} at 3 ppm within T₂ hyperintense regions and the Extended Glasgow Coma Scale (GOSE) at 6 months from the time of injury ($R^2 = 0.5334$, $P = 0.0107$). C) Association between time from injury and MRI exam (log-scale) and MTR_{asym} at 3 ppm within T₂ hyperintense regions ($R^2 = 0.6317$, $P = 0.0004$).

EPI can be used to obtain high resolution, whole brain images within clinically realistic scan times.

It is important to point out a few limitations to the current study. First, no correction for B₁ inhomogeneities were performed. As B₁ can vary as much as ± 20% (Sengupta et al., 2017), our simulations (not shown) suggest that acidic tissue within areas of high amino acid concentration (~50 mM) can have MTR_{asym} at 3 ppm that varies ± 15% (e.g. from 3.65% to 4.3%). Additionally, the results observed in the current study should be taken as preliminary, since the relatively small number of patients included may not represent the strength of trends observed in a larger cohort of patients with different injury profiles. Future studies with a larger number of patients and inclusion of B₁ correction are warranted.

In summary, the current study demonstrates the feasibility and potential clinical value of pH-weighted amine CEST EPI as a high-resolution imaging tool for identifying tissue most at risk for long-term damage due to cerebral acidosis. Results suggest low pH in contusion and pericontusional regions correlate strongly with clinical function and 6-month outcome measures.

Author disclosure statement

Ellingson – Advisory Board – Hoffman La-Roche; Siemens; Nativis; Medcenna; MedQIA; Bristol Meyers Squibb; Imaging Endpoints; Agios. Paid Consultant – Nativis; MedQIA; Siemens; Hoffman La-Roche; Imaging Endpoints; Medcenna; Agios. Grant Funding – Hoffman La-Roche;

Siemens; Agios; Janssen. Dr. Ellingson also holds a patent on this technology (US Patent #15/577,664; International PCT/US2016/034886).

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References

- Bryant, R.G., 1996. The dynamics of water-protein interactions. *Annu. Rev. Biophys. Biomol. Struct.* 25, 29–53.
- Bullock, R., Zauner, A., Myseros, J.S., Marmarou, A., Woodward, J.J., Young, H.F., 1995. Evidence for prolonged release of excitatory amino acids in severe human head trauma. Relationship to clinical events. *Ann. N. Y. Acad. Sci.* 765, 290–297 (discussion 298).
- Bullock, R., Zauner, A., Woodward, J.J., Myseros, J., Choi, S.C., Ward, J.D., Marmarou, A., Young, H.F., 1998. Factors affecting excitatory amino acid release following severe human head injury. *J. Neurosurg.* 89, 507–518.
- Chamoun, R., Suki, D., Gopinath, S.P., Goodman, J.C., Robertson, C., 2010. Role of extracellular glutamate measured by cerebral microdialysis in severe traumatic brain injury. *J. Neurosurg.* 113, 564–570.
- Choi, C., Coupland, N.J., Bhardwaj, P.P., Kalra, S., Casault, C.A., Reid, K., Allen, P.S., 2006. T2 measurement and quantification of glutamate in human brain in vivo. *Magn. Reson. Med.* 56, 971–977.
- Clausen, T., Khaldi, A., Zauner, A., Reinert, M., Doppenberg, E., Menzel, M., Soukup, J., Alves, O.L., Bullock, M.R., 2005. Cerebral acid-base homeostasis after severe traumatic brain injury. *J. Neurosurg.* 103, 597–607.
- Coles, J.P., Minhas, P.S., Fryer, T.D., Smielewski, P., Aigbirihio, F., Donovan, T., Downey, S.P., Williams, G., Chatfield, D., Matthews, J.C., Gupta, A.K., Carpenter, T.A., Clark, J.C., Pickard, J.D., Menon, D.K., 2002. Effect of hyperventilation on cerebral blood flow in traumatic head injury: clinical relevance and monitoring correlates. *Crit. Care Med.* 30, 1950–1959.
- Diringer, M.N., Videen, T.O., Yundt, K., Zazulia, A.R., Aiyagari, V., Dacey Jr., R.G., Grubb, R.L., Powers, W.J., 2002. Regional cerebrovascular and metabolic effects of hyperventilation after severe traumatic brain injury. *J. Neurosurg.* 96, 103–108.
- Faden, A.I., Demediuk, P., Panter, S.S., Vink, R., 1989. The role of excitatory amino acids and NMDA receptors in traumatic brain injury. *Science* 244, 798–800.
- Finner, S.R., Cohen, J., 2001. Severe traumatic brain injury. *Resuscitation* 48, 77–90.
- Folkersma, H., Foster Dingley, J.C., van Berckel, B.N., Rozemuller, A., Boellaard, R., Huisman, M.C., Lammertsma, A.A., Vandertop, W.P., Molthoff, C.F., 2011. Increased cerebral (R)-[(11)C]PK11195 uptake and glutamate release in a rat model of traumatic brain injury: a longitudinal pilot study. *J. Neuroinflammation* 8, 67.
- Ghajar, J., 2000. Traumatic brain injury. *Lancet* 356, 923–929.
- Gupta, A.K., Zygun, D.A., Johnston, A.J., Steiner, L.A., Al-Rawi, P.G., Chatfield, D., Shepherd, E., Kirkpatrick, P.J., Hutchinson, P.J., Menon, D.K., 2004. Extracellular brain pH and outcome following severe traumatic brain injury. *J. Neurotrauma* 21, 678–684.
- Harris, J.L., Yeh, H.W., Choi, I.Y., Lee, P., Berman, N.E., Swerdlow, R.H., Craciunas, S.C., Brooks, W.M., 2012. Altered neurochemical profile after traumatic brain injury: (1)H-MRS biomarkers of pathological mechanisms. *J. Cereb. Blood Flow Metab.* 32, 2122–2134.
- Harris, R.J., Cloughesy, T.F., Liau, L.M., Prins, R.M., Antonios, J.P., Li, D., Yong, W.H., Pope, W.B., Lai, A., Nghiemphu, P.L., Ellingson, B.M., 2015. pH-weighted molecular imaging of gliomas using amine chemical exchange saturation transfer MRI. *Neuro-Oncology* 17, 1514–1524.
- Harris, R.J., Cloughesy, T.F., Liau, L.M., Nghiemphu, P.L., Lai, A., Pope, W.B., Ellingson, B.M., 2016. Simulation, phantom validation, and clinical evaluation of fast pH-weighted molecular imaging using amine chemical exchange saturation transfer echo planar imaging (CEST-EPI) in glioma at 3 T. *NMR Biomed.* 29, 1563–1576.
- Harris, R.J., Yao, J., Chakhoyan, A., Raymond, C., Leu, K., Liau, L.M., Nghiemphu, P.L., Lai, A., Salamon, N., Pope, W.B., Cloughesy, T.F., Ellingson, B.M., 2018. Simultaneous pH-sensitive and oxygen-sensitive MRI of human gliomas at 3 T using multi-echo amine proton chemical exchange saturation transfer spin-and-gradient echo echo-planar imaging (CEST-SAGE-EPI). *Magn. Reson. Med.* 80, 1962–1978.
- Hovda, D.A., Becker, D.P., Katayama, Y., 1992. Secondary injury and acidosis. *J. Neurotrauma* 9 (Suppl. 1), S47–S60.
- Hum, P.D., Traystman, R.J., 1996. pH-associated brain injury in cerebral ischemia and circulatory arrest. *J. Intensive Care Med.* 11, 205–218.
- Jenkinson, M., Bannister, P., Brady, M., Smith, S., 2002. Improved optimization for the robust and accurate linear registration and motion correction of brain images. *Neuroimage* 17, 825–841.
- Jiang, J.-Y., Gao, G.-Y., Li, W.-P., Yu, M.-K., Zhu, C., 2002. Early indicators of prognosis in 846 cases of severe traumatic brain injury. *J. Neurotrauma* 19, 869–874.
- Jin, T., Kim, S.G., 2014. Advantages of chemical exchange-sensitive spin-lock (CESL) over chemical exchange saturation transfer (CEST) for hydroxyl- and amine-water proton exchange studies. *NMR Biomed.* 27, 1313–1324.
- Jin, T., Wang, P., Zong, X., Kim, S.G., 2012. Magnetic resonance imaging of the Amine-Proton EXchange (APEX) dependent contrast. *Neuroimage* 59, 1218–1227.
- Jones, C.K., Huang, A., Xu, J., Edden, R.A., Schar, M., Hua, J., Oskolkov, N., Zaca, D., Zhou, J., McMahon, M.T., Pillai, J.J., van Zijl, P.C., 2013. Nuclear Overhauser enhancement (NOE) imaging in the human brain at 7T. *Neuroimage* 77, 114–124.
- Kalimo, H., Rehncrona, S., Soderfeldt, B., Olsson, Y., Siesjo, B.K., 1981. Brain lactic acidosis and ischemic cell damage: 2. Histopathology. *J. Cereb. Blood Flow Metab.* 1, 313–327.
- Kirkness, C.J., Burr, R.L., Cain, K.C., Newell, D.W., Mitchell, P.H., 2005. Relationship of cerebral perfusion pressure levels to outcome in traumatic brain injury. *Acta Neurochir. Suppl.* 95, 13–16.
- Levin, H.S., Boake, C., Song, J., McCauley, S., Contant, C., Diaz-Marchan, P., Brundage, S., Goodman, H., Kotrla, K.J., 2001. Validity and sensitivity to change of the extended Glasgow Outcome Scale in mild to moderate traumatic brain injury. *J. Neurotrauma* 18, 575–584.
- Li, Y., Srinivasan, R., Ratiney, H., Lu, Y., Chang, S.M., Nelson, S.J., 2008. Comparison of T(1) and T(2) metabolite relaxation times in glioma and normal brain at 3T. *J. Magn. Reson. Imaging* 28, 342–350.
- Liepinsh, E., Otting, G., 1996. Proton exchange rates from amino acid side chains—implications for image contrast. *Magn. Reson. Med.* 35, 30–42.
- Maas, A.I., Stocchetti, N., Bullock, R., 2008. Moderate and severe traumatic brain injury in adults. *Lancet Neurol.* 7, 728–741.
- Marshall, Lawrence F., Gautille, Theresa, Klauber, Melville R., Eisenberg, Howard M., Jane, John A., Luerssen, Thomas G., Marmarou, Anthony, Foulkes, Mary A., 1991. The outcome of severe closed head injury. *Spec. Suppl.* 75, S28–S36.
- McDonald, J.W., Bhattacharyya, T., Sensi, S.L., Lobner, D., Ying, H.S., Canzoniero, L.M., Choi, D.W., 1998. Extracellular acidity potentiates AMPA receptor-mediated cortical neuronal death. *J. Neurosci.* 18, 6290–6299.
- McIntosh, T.K., Faden, A.I., Bendall, M.R., Vink, R., 1987. Traumatic brain injury in the rat: alterations in brain lactate and pH as characterized by 1H and 31P nuclear magnetic resonance. *J. Neurochem.* 49, 1530–1540.
- Mellergard, P., Ouyang, Y.B., Siesjo, B.K., 1994. The regulation of intracellular pH is strongly dependent on extracellular pH in cultured rat astrocytes and neurons. *Acta Neurochir. Suppl. (Wien)* 60, 34–37.
- Miller, J.D., 1985. Head injury and brain ischaemia—implications for therapy. *Br. J. Anaesth.* 57, 120–130.
- Nagy, Z., Szabo, M., Huttner, I., 1985. Blood-brain barrier impairment by low pH buffer perfusion via the internal carotid artery in rat. *Acta Neuropathol.* 68, 160–163.
- Palmer, A.M., Marion, D.W., Botscheller, M.L., Swedlow, P.E., Styren, S.D., DeKosky, S.T., 1993. Traumatic brain injury-induced excitotoxicity assessed in a controlled cortical impact model. *J. Neurochem.* 61, 2015–2024.
- Perry, T.L., Hansen, S., Berry, K., Mok, C., Lesk, D., 1971. Free amino acids and related compounds in biopsies of human brain. *J. Neurochem.* 18, 521–528.
- Prabhakar, H., Sandhu, K., Bhagat, H., Durga, P., Chawla, R., 2014. Current concepts of optimal cerebral perfusion pressure in traumatic brain injury. *J. Anaesthesiol. Clin. Pharmacol.* 30, 318–327.
- Sengupta, A., Gupta, R.K., Singh, A., 2017. Evaluation of B1 inhomogeneity effect on DCE-MRI data analysis of brain tumor patients at 3T. *J. Transl. Med.* 15, 242.
- Shiogai, T., Nara, I., Saruta, K., Hara, M., Saito, I., 1999. Continuous monitoring of cerebrospinal fluid acid-base balance and oxygen metabolism in patients with severe head injury: pathophysiology and treatments for cerebral acidosis and ischemia. *Acta Neurochir. Suppl.* 75, 49–55.
- Siesjo, B.K., Katsura, K., Mellergard, P., Ekholm, A., Lundgren, J., Smith, M.L., 1993. Acidosis-related brain damage. *Prog. Brain Res.* 96, 23–48.
- Sun, P.Z., Lu, J., Wu, Y., Xiao, G., Wu, R., 2013. Evaluation of the dependence of CEST-EPI measurement on repetition time, RF irradiation duty cycle and imaging flip angle for enhanced pH sensitivity. *Phys. Med. Biol.* 58, N229–240.
- Swanson, R.A., Benington, J.H., 1996. Astrocyte glucose metabolism under normal and pathological conditions in vitro. *Dev. Neurosci.* 18, 515–521.
- Swanson, R.A., Farrell, K., Simon, R.P., 1995. Acidosis causes failure of astrocyte glutamate uptake during hypoxia. *J. Cereb. Blood Flow Metab.* 15, 417–424.
- Timofeev, I., Carpenter, K.L., Nortje, J., Al-Rawi, P.G., O'Connell, M.T., Czosnyka, M., Smielewski, P., Pickard, J.D., Menon, D.K., Kirkpatrick, P.J., Gupta, A.K., Hutchinson, P.J., 2011. Cerebral extracellular chemistry and outcome following traumatic brain injury: a microdialysis study of 223 patients. *Brain* 134, 484–494.
- Timofeev, I., Nortje, J., Al-Rawi, P.G., Hutchinson, P.J., Gupta, A.K., 2013. Extracellular brain pH with or without hypoxia is a marker of profound metabolic derangement and increased mortality after traumatic brain injury. *J. Cereb. Blood Flow Metab.* 33, 422–427.
- Traber, F., Block, W., Lamerichs, R., Gieseke, J., Schild, H.H., 2004. 1H metabolite relaxation times at 3.0 tesla: measurements of T1 and T2 values in normal brain and determination of regional differences in transverse relaxation. *J. Magn. Reson. Imaging* 19, 537–545.
- Vespa, P., Prins, M., Ronne-Engstrom, E., Caron, M., Shalmon, E., Hovda, D.A., Martin, N.A., Becker, D.P., 1998. Increase in extracellular glutamate caused by reduced cerebral perfusion pressure and seizures after human traumatic brain injury: a microdialysis study. *J. Neurosurg.* 89, 971–982.
- Vespa, P., Bergsneider, M., Hattori, N., Wu, H.-M., Huang, S.C., Martin, N.A., Glenn, T.C., McArthur, D.L., Hovda, D.A., 2005. Metabolic crisis without brain ischemia is

- common after traumatic brain injury: a combined microdialysis and positron emission tomography study. *J. Cereb. Blood Flow Metab.* 25, 763–774.
- Vink, R., McIntosh, T.K., Weiner, M.W., Faden, A.I., 1987. Effects of traumatic brain injury on cerebral high-energy phosphates and pH: a ^{31}P magnetic resonance spectroscopy study. *J. Cereb. Blood Flow Metab.* 7, 563–571.
- Wang, W., Zhang, H., Lee, D.H., Yu, J., Cheng, T., Hong, M., Jiang, S., Fan, H., Huang, X., Zhou, J., Wang, J., 2017. Using functional and molecular MRI techniques to detect neuroinflammation and neuroprotection after traumatic brain injury. *Brain Behav. Immun.* 64, 344–353.
- Willis, C., Lybrand, S., Bellamy, N., 2004. Excitatory amino acid inhibitors for traumatic brain injury. *Cochrane Database Syst. Rev.* 1, CD003986 (Review. PubMed PMID: 14974051).
- Woessner, D.E., Zhang, S., Merritt, M.E., Sherry, A.D., 2005. Numerical solution of the Bloch equations provides insights into the optimum design of PARACEST agents for MRI. *Magn. Reson. Med.* 53, 790–799.
- Woischneck, D., Firsching, R., 1998. Efficiency of the Glasgow Outcome Scale (GOS)-Score for the long-term follow-up after severe brain injuries. *Acta Neurochir. Suppl.* 71, 138–141.
- Wright, P.J., Mougin, O.E., Totman, J.J., Peters, A.M., Brookes, M.J., Coxon, R., Morris, P.E., Clemence, M., Francis, S.T., Bowtell, R.W., Gowland, P.A., 2008. Water proton T1 measurements in brain tissue at 7, 3, and 1.5 T using IR-EPI, IR-TSE, and MPRAGE: results and optimization. *MAGMA* 21, 121–130.
- Xu, S., Zhuo, J., Racz, J., Shi, D., Roys, S., Fiskum, G., Gullapalli, R., 2011. Early microstructural and metabolic changes following controlled cortical impact injury in rat: a magnetic resonance imaging and spectroscopy study. *J. Neurotrauma* 28, 2091–2102.
- Yao, J., Ruan, D., Raymond, C., Liao, L.M., Salamon, N., Pope, W.B., Nghiemphu, P.L., Lai, A., Cloughesy, T.F., Ellingson, B.M., 2018 Sep. Improving B(0) correction for pH-weighted Amine Proton Chemical Exchange Saturation Transfer (CEST) imaging by Use of K-means clustering and Lorentzian estimation. *Tomography* 4 (3), 123–137. <https://doi.org/10.18383/j.tom.2018.00017>. (PubMed PMID: 30320212; PubMed Central PMCID: PMC6173788).
- Yermolaieva, O., Leonard, A.S., Schnitzler, M.K., Abboud, F.M., Welsh, M.J., 2004. Extracellular acidosis increases neuronal cell calcium by activating acid-sensing ion channel 1a. *Proc. Natl. Acad. Sci. U. S. A.* 101, 6752–6757.
- Ying, W., Han, S.K., Miller, J.W., Swanson, R.A., 1999. Acidosis potentiates oxidative neuronal death by multiple mechanisms. *J. Neurochem.* 73, 1549–1556.
- Yokota, H., Yamamoto, Y., Naoe, Y., Fuse, A., Sato, H., Unemoto, K., Kurokawa, A., 2000. Measurements of cortical cellular pH by intracranial tonometer in severe head injury. *Crit. Care Med.* 28, 3275–3280.
- Zauner, A., Clausen, T., Alves, O.L., Rice, A., Levasseur, J., Young, H.F., Bullock, R., 2002. Cerebral metabolism after fluid-percussion injury and hypoxia in a feline model. *J. Neurosurg.* 97, 643–649.